



Determination of Phytochemical Compositions of Leaves and Flowers of *Cassia auriculata*

K. Parani ^{a*}

^a Department of Botany, Sri Parasakthi College for Women (Autonomous), Courtallam, Tenkasi (Dt.), (Manonmaniam Sundaranar University, Abishekapatti, Tirunelveli) Tamil Nadu, India.

Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

Article Information

DOI: <https://doi.org/10.9734/ejmp/2024/v35i61204>

Open Peer Review History:
This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/118498>

Original Research Article

Received: 20/04/2024
Accepted: 22/06/2024
Published: 16/08/2024

ABSTRACT

The present study reveals the morphological characteristics, organoleptic and fluorescence analysis by using various chemicals and reagents to examine the presence of phytochemicals visually in powdered samples of leaves and flowers of *Cassia auriculata*. The qualitative analysis represents the presence of various phytochemicals viz., steroids, reducing sugars, sugars, alkaloids, phenols, flavonoids, saponins, tannins, Anthraquinones and amino acids in crude extracts (aqueous, ethanol and petroleum ether) of leaves and flowers of *Cassia auriculata*. The presence of minerals i.e., potassium, phosphorous, sulphur, calcium, were observed in leaves and flowers of *Cassia auriculata* by qualitatively. Quantitative analysis revealed maximum amount of flavonoids in flowers of *Cassia auriculata* and lesser amounts of alkaloids were recorded in leaves and flowers of *Cassia* respectively.

Keywords: *Cassia auriculata*; fluorescence analysis; phytochemicals.

*Corresponding author: E-mail: barani12101975@gmail.com;

Cite as: Parani, K. 2024. "Determination of Phytochemical Compositions of Leaves and Flowers of *Cassia Auriculata*". *European Journal of Medicinal Plants* 35 (6):13-20. <https://doi.org/10.9734/ejmp/2024/v35i61204>.

1. INTRODUCTION

“Phytochemicals are a large group of plant-derived compounds that are hypothesized to be responsible for much of the disease reduction conferred by a diet high in fruits vegetables, beans, cereal and plant-based beverages such as tea and wine. Based on their chemical structure phytochemicals can be grouped into such groups as tannins, flavonoids, glycosides, saponins, alkaloids, triterpenoids and sterols” [1].

“For a long period of time, plants have been a valuable source of natural products for maintaining human health, especially in the last decade, with more intensive studies for natural therapies. About 80% of individuals from developed countries use traditional medicine, which has compounds derived from medicinal plants. Sources for new safe, biodegradable and renewable drugs. The use of plants as therapeutic agents in addition to being used as food is age-long” [2].

“Though the therapeutic uses of plants by the primitive people lack scientific explanations there is a great awareness in the use and significance of these medicinal floras by the World Health Organization in several resource-poor nations” [3,4]. “This has led to intensified efforts in the documentation of medicinal plants” [5].

“Therefore, such plants should be investigated to better understand their properties, safety and efficiency” [6]. The cost of drugs in use today is too expensive for the majority of the population in the third world countries and therefore the search for some cheap sources of antimicrobial substances in nature become inevitable plants are good.

Cassia auriculata Linn (Family: Caesalpinaceae) distributed throughout hot deciduous forests of India and holds a very prestigious position in Ayurveda and Siddha system of medicine. The plant has been reported to possess antipyretic [7]; hepato protective [8]; anti diabetic, antiperoxidative and antihyperglycemic [9] and microbicidal activity [10]. “The flowers are used to treat urinary discharges, nocturnal emissions, diabetes and throat irritation” [11]. Hence, the objectives of the present study is focused on evaluating the phytochemicals qualitatively and quantitatively of the *Cassia auriculata* using various extracts.

2. MATERIALS AND METHODS

2.1 Collection of Plant Materials

Leaves and flowers of *Cassia auriculata* (Caesalpinaceae) were collected from Karuppanathi Dam near Chokampatti, Tenkasi (TK.) Tamil Nadu.

2.2 Organoleptic Study

The plant powder characteristics such as color, odor, taste and nature were evaluated.

2.3 Preparation of Crude extracts

The collected plant samples were thoroughly washed under running tap water and shade dried. The samples were pulverised with the help of a blender / mixer and soaked in aqueous, ethanol and petroleum ether were prepared by macerating one gram of powder with 10ml of solvents taken in flasks wrapped separately in Erlenmeyer flasks. The preparation were allowed to stand for 4 hrs. at room temperature. Then the extracts were filtered using Whatmann filter No. 1 and stored for further use. The crude extracts were analysed qualitatively and quantitatively and percentage yield of the extract was determined by using the equation [12].

$$\text{Yield (\%)} = \frac{W_2 - W_1}{W_0} \times 100$$

Where W_2 - weight of the extract and container
 W_1 - weight of the empty container and W_0 is the weight of the initial dried sample.

2.4 Fluorescence Analysis of the Powder

The fluorescence analysis of powdered samples i.e., leaves and flower mixed with different solvents and reagents were carried out using long ultraviolet (UV) lamps (365nm) and visible wavelengths [13 – 15].

2.5 Preliminary Phytochemical Analysis

The qualitative tests for extracts to detect the presence of phytochemicals such as alkaloid, tannin, saponin, flavonoid and phenol were carried out using standard procedures [16].

2.6 Quantitative analysis of Phytochemicals

2.6.1 Phenol determination

100 mg of the extract of the sample was weighed accurately and dissolved in 100 ml of triple distilled water (TDW). 1 ml of this solution was

transferred to a test tube, then 0.5 ml 2N of the Folin Ciocalteu reagent and 1.5ml 20% of Na₂CO₃ solution was added and ultimately the volume was made up to 8 ml with TDW followed by vigorous shaking and finally allowed to stand for 2 hours after which the absorbance was taken at 765 nm. These data were used to estimate the total phenolic content using a standard calibration curve obtained from various diluted concentrations of gallic acid.

2.6.2 Alkaloid determination

“Five gram of the powdered plant samples were weighed into 200 ml of 20% acetic acid in ethanol was added and covered to stand for 4 h. This was filtered and the extract was concentrated using a water bath to one-quarter of the original volume. Concentrated ammonium hydroxide was added drop wise to the extract until the precipitation was complete. The whole solution was allowed to settle and the precipitate was collected by filtration and weighed” [17].

2.6.3 Flavonoid determination

“To estimate flavonoids quantitatively, 10 g of powdered sample of each plant material was extracted twice with 10 ml of 80% aqueous methanol at room temperature. The whole solution was filtered through Whatmann filter paper No.1, the filtrate was later transferred into crucibles, and evaporated to dryness on a water bath to a constant weight” [17].

2.6.4 Tannin determination

“Distilled water (50 ml) was added to 500 mg of the sample taken in a 500 ml flask and kept in a shaker for 1 h. It was filtered into a 50 ml volumetric flask and made up to the mark. Then 5 ml of the filtrate was pipetted out into a test tube and mixed with 2 ml (10 fold diluted) of 0.1 M FeCl₃ in 0.1 N HCl and 0.008 M potassium ferrocyanide. The absorbance was measured at 605 nm within 10min” [18].

3. RESULTS AND DISCUSSION

“Plant based antibacterial preparations are known to have enormous therapeutic potential due to the presence of several antibacterial substances” [19]. “In order to identify the antibacterial active compounds of the herbs or medicinal plants, such factor should be taken into consideration including the extraction and bio assay techniques employed. Generally, the type

of solvent used for the extraction plays a significant role in the solubility of the active principles of plant material that not only affected the amount of representative compounds where consequently will influence the antibacterial activity of the extract” [20]. Flowers of *Cassia auriculata* are showy with dense bunches, brightly yellow color corolla, leaf obovate with tapering apex, dark or prominent vein, waxy coated and coriaceous (Table 1).

3.1 Organoleptic Studies

“It is an important parameter of powder analysis which is technique for the qualitative detection of morphological and sensory profile of drugs” [[21]. The study revealed the characteristic color, odor, taste and nature of powdered medicinal species (Table 2). “The results of pharmacognostical and phytochemical studies conducted in the bark and leaves of *Terminalia travancorensis* Wight & Arn. (Combretaceae), a tree, endemic to the Western Ghats and their pharmacognostical studies included the organoleptic, physico-chemical and fluorescence analysis of the bark and leaf powder” [22].

3.2 Percentage Yield of Crude Extracts

Table 3 represents the percentage yield of crude extracts in medicinal plants in which high yield occurred in ethanolic extract of leaves (24%) and low yield was observed in flowers of *Cassia auriculata* (6%). “Highest yield percentage accounted for 1.70% was obtained in maceration with methanol followed by ethyl acetate at 1.28% and n-hexane at 0.93% in a study done on *Psidium guajava* leaves extract, suggested that methanol was the best solvent for solubility of several compounds” [23]. “Nevertheless, the preferred extraction method should be simple, fast, economical and importantly able to retain the important phytoconstituents” [24].

3.3 Fluorescence Analysis

In fluorescence analysis study shows specific colour appeared with specific reagents (Table 4). The powdered leaves of *Cassia* recorded as light green to brown colour under ordinary white light and fluorescent green to dark brown appeared in long UV light (365 nm). The *Cassia* flower powder appeared as light yellow to dark black in ordinary light, while under long UV light (365 nm) exposed showed fluorescence green to dark red in colour.

Table 1. Morphological features of *Cassia auriculata*

S. No	Parameters	Leaf	Flower
1.	Habit	Shrub	Shrub
2.	Root system	Tap root	Tap root
3.	Stem	Branches	Branches
4.	Leaf	Lanceolate	Lanceolate
5.	Flower	Large and showy	Large and showy
6.	Colour	Green	Yellow
7.	Inflorescence	Racemose	Racemose

Table 2. Organoleptic character of *Cassia auriculata*

Characters	Leaf	Flower
Odour	No smell	No smell
Taste	Bitter	Bitter
Colour	Normal green	Yellow
Texture	Fine	Fine

Table 3. Percentage yield of *Cassia auriculata*

Ethanolic Extract	Yield percentage			
	Wt. of initial dried sample (W_0)	Wt. of empty container (W_1)	Wt. of extract & container (W_2)	Yield (%)
Leaves	1.0	65.40	65.64	24
Flowers	1.0	61.37	61.43	6

Table 4. Fluorescence analysis of *Cassia auriculata*

S. No	Reagent Used	Cassia leaf		Cassia flower	
		Long UV light (365 nm)	Visible light	Long UV light (365 nm)	Visible light
1.	Concentrated HNO_3	Dark brown	Light brown	Dark red	Dark red
2.	Concentrated HCL	Greenish brown	Pale green	Dark green	Light green
3.	Acetone	Dark brown	Brown	Light yellow	Light yellow
4.	NH_3 + Ammonia	Brownish green	Brown	Flourescence green	Light yellow
5.	Chloroform	Dark green	Light green	Dark yellow	Light yellow
6.	Benzene	Dark green	Light green	Greenish yellow	Light yellow
7.	Ethanol	Dark brown	Light brown	Greenish yellow	Light yellow
8.	Petroleum ether	Dark green	Light green	Dark yellow	Light yellow
9.	Glacial acidic acid	Dark brown	Dark brown	Greenish yellow	Light yellow
10.	HNO_3 + NH_3	Greenish brown	Brown	Dark black	Dark red
11.	H_2SO_4	Dark brown	Dark brown	Dark black	Dark black
12.	50% HNO_3	Greenish brown	Brown	Dark black	Dark red
13.	50% HCL	Dark green	Light green	Greenish yellow	Light yellow
14.	1N Aqueous NaOH	Dark green	Light green	Dark red	Dark black
15.	1N Alcoholic NaOH	Dark brown	Dark brown	Dark black	Light black
16.	50% H_2SO_4	Dark brown	Light brown	Brownish green	Light yellow
17.	Ferric chloride	Flourescent green	Brown	Dark red	Light red
18.	40% of NaOH + 10% Lead acetate	Dark green	Brown	Greenish yellow	Light red

Table 5. Phytochemical analysis of *Cassia auriculata* in various extracts

S.No	Phytoconstituents	Aqueous		Ethanollic		Petroleum ether	
		Leaf	Flowers	Leaf	Flowers	Leaf	Flower
1	Steroids	-	-	+	-	+	-
2	Reducing sugar	-	+	-	+	-	-
3	Sugar	+	+	-	+	+	+
4	Alkaloids	+	-	-	-	+	-
5	Phenol	+	+	-	+	+	+
6	Flavonoids	+	-	+	+	-	-
7	Saponin	+	-	+	+	-	+
8	Tannin	+	+	-	+	+	+
9	Anthroquinine	-	-	-	-	-	-
10	Amino acids	+	-	-	+	-	-

(+) : Present (-): Absent

Table 6. Quantitative analysis of phytochemicals

S.NO	Phytoconstituents	<i>Cassia</i> leaf	<i>Cassia</i> flower
1.	Flavonoids (%)	14	28
2.	Phenols (mg / g)	0.16	0.20
3.	Tannins (mg / g)	0.65	0.47
4.	Alkaloids (%)	6.0	5.0

“Herbal drug which are used in various traditional medicine needs detailed investigation with an ethano pharmacological approach. The present study provides information in respect of the identification, standardization of herbal drug of *Cassia auriculata* of Ayurvedic compendia. Correct identification and quality assurance of the starting materials is an essential pre requisite to ensure reproducible quality of herbal medicine which will contribute to its safety and efficacy” [25].

“The fluorescence analysis is adequately sensitive and enables the precise and accurate determination over a satisfactory concentration range without several time consuming dilution steps prior to analysis of pharmaceutical samples” [26]. The fluorescence colour is specific for each compound. A non fluorescent compound may fluorescence it mixed with impurities that are fluorescent.

3.4 Qualitative Analysis

As shown in the Table 5. maximum number of phytochemicals were observed in water extract of *Cassia* leaf followed by ethanol extracts of *Cassia* flower. Lesser number of phytochemicals was seen in ethanol extract of *Cassia* leaves.

“Our results are positively correlated with ethanolic extracts revealing the presence of a high concentration of tannins, reducing sugars

and steroids in the stem, bark and roots” [27]. Flavonoids, phenolics and protein were prevented in high concentration in the stem, bark while anthraquinone, glycosides and alkaloids were present in the leaves and roots of *Cassia abbreviate* respectively.

“Bioactivity properties of herbs are where closely related to their phytochemicals constituents which are classified into various major groups” [28]. Ethanolic extract of *Cassia* flower possess reducing sugars, phenol, tannins, and steroids can summarized in Table 3. However, it is important to highlight that the type of diluent used was the main factor that could influence in variation of phytoconstituents being extracted.

“While in aqueous extract of *Cassia* flower showed the absence of phytoconstituents namely alkaloids, saponins, and antheroquinone other study that evaluated the existence of phytochemicals of petroleum ether, ethanol and aqueous extracts also revealed the difference in the solubility of active compounds” [29].

“Phenolic compounds were the most common secondary metabolites implicated with microbial growth inhibitory action in herbs” [30,31]. Plants are rich in wide variety of secondary metabolites such as tannin, terpenoids, alkaloids and flavonoids etc., which have been *in vitro* to have antibacterial and antifungal properties.

3.5 Quantitative Estimation of Major Phyto Components

Table 6 reveals the amount of phytochemicals quantitatively, in which more amount of flavonoids was recorded in flowers of *Cassia auriculata* (28%) followed by leaves of *Cassia* (14%). Lowest amount of alkaloids was observed in leaves and flowers of *Cassia* (6% and 5% of alkaloids) respectively, while tannins were more in leaves of *Cassia* (0.65 mg/g) followed by *Cassia* flowers (0.47 mg/g).

“Plant based compounds have several biological applications. An alkaloid compound has been reported to exhibit lethal effects against colon and breast cancer cells and has been used for antimicrobial, antiviral, antiprotozoal and anti tumor applications” [32]. “Flavonoids have been used for anti diabetic, anti microbial activities, anti -inflammatory and anti aging preparations” [33]. “Previous researchers have shown that the plant phenolic compounds offer the role of potential natural antioxidants” [34,35,36,37].

“Flavonoids and phenols have raised particular interest because of their potential biological characteristics as antioxidant, antiestrogenic, anti inflammatory, immune modulatory, cardio protective and anti carcinogenic compounds” [38,39]. “Tannins play an essential role in many biological applications because of their anti inflammatory, cardio protective and anti microbial properties” [40]. “Presence of alkaloids, tannins, total phenols, carbohydrates, total tannins, saponins, terpenoids and total glycosides in varying content using various solvents viz., ethanol, methanol, acetone, chloroform, petroleum ether and also in water, clearly shown that the more number of photochemical compounds are maximum soluble in ethanol solvent” [41].

3.6 Qualitative Analysis of Minerals

The presence of minerals i.e., potassium, phosphorous, sulphur, calcium, were observed in leaves and flowers of *Cassia auriculata* by qualitatively. Similarly reports that different plant species, elemental accumulation depends on various factors such as the type of soil, fertilization method, plant species and environmental conditions [42].

Proximate and mineral nutrient analysis validates the significance of the extracts with a high amount of carbohydrates and proteins along with significantly high amount of zinc, iron,

manganese, calcium, magnesium and potassium involved in various metabolic reactions of *Calligomumcrinitum* [43].

4. CONCLUSION

The present study reveals the morphological characteristics, organoleptic and fluorescence analysis by using various chemicals and reagents to examine the presence of phytochemicals visually in powdered samples of leaves and flowers of *Cassia auriculata*. The presence of minerals i.e., potassium, phosphorous, sulphur, calcium, were observed in leaves and flowers of *Cassia auriculata* by qualitatively. The maximum amount of flavonoids was found in flowers of *Cassia auriculata* and lesser amounts of alkaloids were found in leaves and flowers of *Cassia* respectively.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Tiwari P, Kumar B, Kaur M, Karu G, Karu H. Phytochemical screening and extraction: A review. International Pharmacia Scientia. 2011;1:1.
2. Sachin Chaudhary, Anit Kumar. Phytochemical analysis and assessment of *In vitro* antihelminthic activity of *Cassia auriculata* Linn. Leaves; 2014.
3. Dutta AC. Botany for degree students. Oxford University Press, London. 1994; 73.
4. WHO. WHO traditional medicine strategy 2002- 2005. WHO, Geneva; 2002.
5. Perumal SR, Ignacimuthu S. Antibacterial activity of some folklore medicinal plants

- used by tribes in Western Ghats of India. J. Ethnopharmacol. 2000;69:63-71.
6. Muthukumar P, Elayarani M, Shanmuganathan P, Cholarajan A. Antimicrobial Activities of *Cassia auriculata* L. and *Morinda tinctoria* Roxb. International Journal of Research in Pure and Applied Microbiology. 2001;1(2):9-12.
 7. Wealth of India. Raw materials. Publications and Information Directorate, Council of Scientific and Industrial Research, New Delhi. 1950;2:95.
 8. Rao KN, Vedavathy S. Antipyretic activity of six indigenous medicinal plants of Tirmula hills. J. Ethnopharmacol. 1991; 33:193-196.
 9. Manickam P, Namasivaqyam N, Periyasamy V, Rajagopal S. Effect of *Cassia auriculata* leaf extract on lipids in rats with alcoholic liver injury. Asia Pacific J. Cli. Nut. 2002;11:57-163.
 10. Pari L, Latha M. Antihyperglycaemic effect of *Cassia auriculata* in experimental diabetes and its effects on key metabolic enzymes involved in carbohydrate metabolism. Cli. Exp. Pharmacol. Physiol. 2003;30:38-43.
 11. Prakash SK. Effects of Herbal extracts towards microbicidal activity against pathogenic *Escherichia coli* in Poultry. Int. J. Poultry Sci. 2006;5:259-261.
 12. Anokwuru CP, Anyasor GN, Ajibaye O, Fakoya O, Okebugwu P. Effect of extraction solvents on phenolic, flavonoid and antioxidant activities of three Nigerian Medicinal plants. Nat. Sci. 2011;9: 53 - 61.
 13. Chase CR, Pratt RS. Fluorescence of powdered vegetable drugs with particular reference to development of a system of identification. J. Am. Pharmacol. Assoc. 1949;38:32.
 14. Kokoshi CJ, Kokoshi RJ, Sharma FT. Fluorescence of powdered vegetable drugs under Ultraviolet radiation. J. Pharm. Asses. 1958;47:715 - 717
 15. Wilson Color Chart – Horticultural Color Chart,. Vol. 1,2, Henry Stone and Son Ltd. Banbury, Great Britain, 1938, 1941, 1 - 100, 10 - 200.
 16. Harborne JB. Phytochemical Methods, A guide to modern technique of plant analysis, Chapman and Hall, London. 1998;108 - 148.
 17. Kumaran A, Karunakaran JR. Antioxidant Activities of the Methanol Extract of *Cardiospermum halicacabum*. Pharmaceutical Biology. 2006;44(2):146 – 151.
 18. Van-Burden TP, Robinson T. The biochemistry of alkaloids. Springer, Heidelberg, New York; 1981
 19. Srinivasan D, Nathan S, Suresh T, Perumalsamy PL. Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. J. Ethnopharmacol. 2001;74:217-220.
 20. Nair R, Kalariya T, Chanda S. Antibacterial activity of some selected Indian medicinal flora. Turk. J. Biol. 2005;29:41- 47.
 21. Kokate CK, Purohit AP, Gokhale SB. Text book of Pharmacognosy. 18th ed. Pune: Nirali Prakashan; 2002.
 22. Lakshmi M, Bindu RN, Chandrasekhara Pillai PK. Pharmacognostic evaluation and phytochemical analysis of bark and leaves of *Terminalia travancorensis* Wight and Arn. (Combretaceae). Journal of Pharmacy Research. 2012;5(4):1988 – 1991.
 23. Shafiei SNS. *In-vitro* antibacterial activity and phytochemical screening of bioactive compounds from Guava (*Psidium guajava* L.) crude leaf extracts. M.Sc. Thesis, Universiti Putra Malaysia, Malaysia; 2012.
 24. Annegowda HV, Tan PV, Mordi MN, Ramanathan S, Hamdan MR, Sulaiman MH, Mansor SM. TLC – bioautography-guided isolation, HPTLC and GC-MS assisted analysis of bioactives of *Piper betle* leaves extract obtained from various extraction techniques: *In vitro* evaluation of phenolic content, antioxidant and antimicrobial activities. Food Anal. Methods. 2013;6:715 - 726.
 25. Ghildiyal S, Gautam MK, Joshi VK, Goel RK. Pharmacognostical study of *Hedychium spicatum* (Ham – Ex-n Smith) rhizome. Asian Journal of Tropical Biomedicine, (In press); 2012.
 26. Pimenta AM, Montenegro MC, Ara-Ujo AN, Martinez JC. Application of sequential injections analysis to pharmaceutical analysis. Journal of Pharmaceutical Biomedical Analysis. 2006;40:16-34.
 27. Huang Q, Liu X, Zhao G, Hu T, Wang Y. Potential and challenges of tannins as an alternative to in-feed antibiotics for farm animal production. Anim. Nutr. 2018; 4(2):137–150.
 28. Al-Daihan S, Al-Faham M, Al-Shawi N, Almayman R, Brnawi A, Zargar S, Bhat RS. Antibacterial activity and phytochemical screening of some medicinal plants commonly used in Saudi

- Arabia against selected pathogenic microorganisms. J. King Saud Univ. Sci. 2013;25:115-120.
29. Syahidah A, Saad CR, Daud HM, Abdelhadi YM. Status and potential of herbal applications in aquaculture: A review. Iran. J. Fish. Sci. 2015;14:27-44.
30. Burt S. Essential oils: Their antibacterial properties and potential applications in foods: A review. Int. J. Food Microbiol. 2004;94:223-253.
31. Witkowska AM, Hickey DK, Alonso-Gomez M, Wilkinson M. Evaluation of antimicrobial activities of commercial herb and spice extracts against selected food-borne bacteria. J. Food Res. 2013;2:37-54.
32. Rinaldi MVN, Diaz IEC, Suffredini IB, Moreno PRH. Alkaloids and biological activity of beriba (*Annonan hypoglauca*). Rev. Bras. Pharmacogn. 2017;27(1):77 - 83.
33. Wang TY, Li Q, Bi KS. Bioactive flavonoids in medicinal plants: Structure, activity and biological fate. Asian J. Pharm. Sci. 2018;13(1):12–23.
34. Dhiman A, Nanda S, Ahmad S. A quest for staunch effects of flavonoids, Utopian protection against hepatic ailments. Arab. J. Chem. 2016;9:1813-23.
35. Takaidza S, Mtunzi F, Pillay M. Analysis of the phytochemical contents and antioxidant activities of crude extracts from Tulbaghia species. J. Tradit. Chin. Med. 2018;38(2):272 – 279.
36. Dermane A, Kporvie AKG, Kindji KP, Metowogo K, Eklug-Gadegboku K. Immunomodulatory and anti-inflammatory activities of hydro-ethanolic extract of *Securidaca longi pedunculata* Fresen leaves. J Herbmed Pharmacol. 2024;13(2): 280-288. DOI: 10.34172/jhp.2024.49352. (For Phenol and alkaloids determination)
37. Widya Twiny Rizki, Winika Sri, Ririn Depita Sari, Siti Marwah Lestari, Rahmadevi Rahmadevi. Tannin extraction from bark of *Cinnamomum burmannii* and its application for use as natural dye and as antioxidant. Indonesian Journal of Fundamental and Applied Chemistry. (For tannin determination). 2024;9(1):35-40.
38. Kumar and Baskar. Screening and quantification of phytochemicals in the leaves and flowers of *Tabernaemontana heyneana* wall-a near threatened medicinal plant. Indian Journal of Natural Product Resources. 2015;5:237 - 243.
39. Tanor EB, Matamane RP, Hapazari I, Magama S. Phytochemical Screening and Antioxidant Analysis of the Ethanolic Extract of Rosehip Seed Press Cake. Curr. J. Appl. Sci. Technol. 2020; 39(35):57-6. Available:<https://journalcjust.com/index.php/CJAST/article/view/3011>
40. Abu Zarin M, Wan HY, Isha A, Armania N. Antioxidant, antimicrobial and cytotoxic potential of condensed tannins from *Leucaena leucocephala* hybrid-Rendang. Food Sci. Human Wellness. 2016;5(2):65 – 75.
41. Suman Kumar Ratnampally, Venkateshwar Chinna. Quantitative analysis of phytochemicals in the Bark extracts of medicinally important plant *Cassia fistula* Linn. Int. J. Curr. Microbiol App, Sci. 2017;6(4):1073 - 1079.
42. Bengtsson H, Oborn I, Jonsson S, Nilsson I, Andersson A. Field balance of some mineral nutrients and trace elements in organic and conventional dairy farming a case study at Objebyn, Sweden. Eur. J. Agron. 2003;20:101-116.
43. Naqbi KMAA, Karthiswaran K, Kurup SS, Abdul Muhsen Alyafei M, Jaleel A. Phytochemicals, proximate composition, mineral analysis and *In vitro* antioxidant activity of *Calligomum crinitum* Boiss. Horticulture. 2022;81(156):1-14.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/118498>